	EALTH LABORATORY N FORM - <u>SLAUGHTER ANIMALS</u> 1105-001 Lab use only					
SAMPLES TAKEN: Date: / / (yyyy/mm/dd) Time of day: SUBMITTED BY: Uveterinarian Owner Agent BILL: 14105-001	Date sent / / (yyyy/mm/dd)					
dized by the Government of Ontario. By submitting samples for testing to AHL, the Submitt	I for testing to the Animal Health Lab ("AHL"). Agricultural animal testing carried out through AHL is subsi- tter acknowledges that s/he is the owner or is a duly authorized agent of the owner. The submitter leems necessary for the purposes of relevant legislation regarding reportable or notifiable diseases and					
AHL USE TEMPLATE 4105-001 (On-farm & slaughter)	Owner Unique ID (max. 40 characters)					
Clinic No 1017740 OMAFRA - CWD	Address					
Address 1 Stone Rd. Guelph ON N1G 4Y2	Premises ID ON Barn Postal Code					
Phone 519-400-5621 Fax 519-826-4375	Phone					
Veterinarian Alexandra Reid Alexandra.Reid@ontario.ca	Farm Barn/Pen/Floor/Batch ID					
Andrew.Vince@ontario.ca, Jeff.perkins@ontario.ca, Manjusri.Wijekoon@ontario.ca,ocvo-reportable-notifiable@ontario.ca	List either Fax or Email (not both) to identify how you want to receive the report from AHL Fax or Email:					
The shaded information must be on final AHL report.						
Animal/ Carcass: On Farm Death Escaped Slaughter Animals HCP = Health Certification Program						
Abattoir name, OMAFRA abattoir number & location:						
Carcass(es) from Healthy Slaughter Animals (HSA) are being held Tissues from HSA have been collected by a vet, government staff Tissues from HSA have been transported to AHL by a vet, govern	for AHL. Yes No					
Species: (check) Note to Submitter: A separate submission form (AHL case #) mus	st be provided for each cervid species					
White-tailed deer Elk Red Deer Elk/Red Deer Hybrid Fallow Deer Reindeer Other Cervids						
Animal Age: <i>years</i> Note to Submitter: All animals must be over 12 months old.						
Samples for CWD: (check) Fresh Tissues Frozen Tissues						
# of Samples: Note to Submitter: Brain stem (obex) & Retropharyngeal (Lymph	node) are required from all species.					
# of heads # of brain stems (obex)	# of brain stems (obex) # of retropharyngeal lymph nodes (RPLN)					
Rabies suspects?	becimen #s					
TB suspects?	becimen #s					
Note: Specimens submitted to the University of Guelph, including carcasses, tio other arrangements are made in writing at the time of submission.	issues and agents isolated from specimens, become the property of the University unless					
Email: ahlinfo@uoguelph.ca Website: http://ahl.uoguelph.ca AHL Guelph: Tel: 519 824-4120 ext: 54530 Fax: 519 827-0961 Guelph, ON M	NW Corner Gordon/McGilvray Mail Bag 2005					
FOR LABORATORY USE ONLY Specimens received b	by: Via: □Courier □Drop-off □Other					

How to Submit SLAUGHTER Samples to AHL for Ontario CWD Voluntary Surveillance Project in Farmed Cervids

- ⇒ An AHL submission form must be completed for all samples to be tested. All samples submitted on a submission form must be processed and reported together.
- ⇒ Hand deliver or courier by Purolator fresh/frozen samples on ice packs. Do not ship perishable samples to arrive on a holiday.
- ⇒ AHL will accept Purolator courier shipments of tissue specimens/heads on an "incoming collect" basis (free to shipper)
 - Contact AHL at 519-824-4120 ext 54530, or by email at ahl.supplies@uoguelph.ca to request a Purolator waybill. Waybills can be emailed directly to the shipper.
- \Rightarrow If submitting heads, they should be frozen prior to shipping, so they are in best condition possible upon arrival at AHL in the event of delays.
- \Rightarrow Heads should be double or triple bagged to prevent leakage & packed on ice
- \Rightarrow Samples should be shipped early in the week (i.e. Monday) so they arrive before the end of the week
- \Rightarrow If submitting intact heads please identify (i.e. 1H, 2H,...).
- \Rightarrow If submitting brain stems (obex) and retropharyngeal lymph nodes (RPLN), both obex and RPLN must be submitted with appropriate identification to AHL fresh on ice packs if possible, otherwise frozen.
- ⇒ Label two specimen whirlpak bags/containers per animal, one for the brain stem/obex (B) and one for the two retropharyngeal lymph nodes (R). Label with initials of producer name or farm name, followed by sample #, followed by tissue type (B or R). Examples of the final labelling for the brain stem/obex and retropharyngeal lymph nodes from animal #1, owned by John Smith are JS1B & JS1R. Place all brain/obex specimens in individual containers into one labelled bag/container & all lymph node specimens in individual containers into another separate labelled bag/container.
- ⇒ All forms of animal identification must be listed in order to be tested. For on-farm deaths, the priority for ID should be 1st CFIA HofA metal ear tag, 2nd farm dangle ear tag.

Labelling of Specimen Containers	Species	Animal Specimen Type	OMAFRA Yellow Held Tag Sticker		
(Use following labelling system - Initials of producer, followed by the sample #, followed by initial of tissue type) Tissue Types: B = Brain Stem (obex), R = Retropharyngeal Lymph Node & H = Head	Elk, red deer, elk-red deer hybrid, fallow deer, white tailed deer, reindeer, other cervids (please specify)	HSA = healthy slaughter animal	<pre># from Abattoir (8numbers)</pre>	CFIA Health of Animals (HofA) Metal Ear Tag # (5 numbers, 1 letter, 1 number)	
			Tissue Types: B = Brain Stem (obex) Use (8 digits including "07")		Farm Dangle Ear Tag Colour & #
		OFD = on-farm death	R = Retropharyngeal Lymph Node & H = Head Use (8 digits including "08")		
		HCP = Herd Certification Program			
The following are 2 examples:					
I. Example JS1B	Elk	HSA	204603 07	85423W2	Yellow W34
2. Example JS1R	Elk	HSA	204603 08	ű	ű
3. Example JS2B	Fallow Deer	CSA	204604 07	85424W2	Yellow W22
I. Example JS2R	Fallow Deer	CSA	204604 08	"	"